

Tom Venema¹, Hinke Dekter², Marijo Parčina³, René Parchen¹

1 BiosparQ, Galileiweg 8, 2333 BD, Leiden, The Netherlands

2 TOPlab, Research Department of Innovative Molecular Diagnostics, University of Applied Sciences Leiden

3 Institute of Medical Microbiology, Immunology and Parasitology, University Hospital Bonn

Introduction

Diagnosis of prosthetic joint infection (PJI) still remains a serious clinical challenge, whereas proper diagnosis is crucial in selecting the right treatment strategy. Loosening of an implant can be caused by both infectious causes as non-infectious causes, e.g. mechanical problems. In non-infectious cases, replacement of the implant can be performed during the same operation (one-stage surgery). In case of infection the patient will be extensively treated with antibiotics for up to several months before a new implant is placed in a second operation (two-stage surgery). Rapid assessment of the presence or absence of infection thus has major consequences for the patient.

Single-cell MALDI-TOF (SC-MALDI-TOF) is a newly developed platform, able to identify strains without previous culturing. This technique is capable of presenting bacterial cells individually to the ionization unit of the mass-spectrometer. Each cell produces a classifiable mass spectrum, enabling a quantitative analysis of a contaminated sample containing a mixture of bacterial species. This strategy provides the opportunity to analyse clinical samples without culturing.

This study investigates the feasibility of SC-MALDI-TOF as a fast PJI diagnostic, using direct aliquots of sonication fluids collected during surgery.

Method

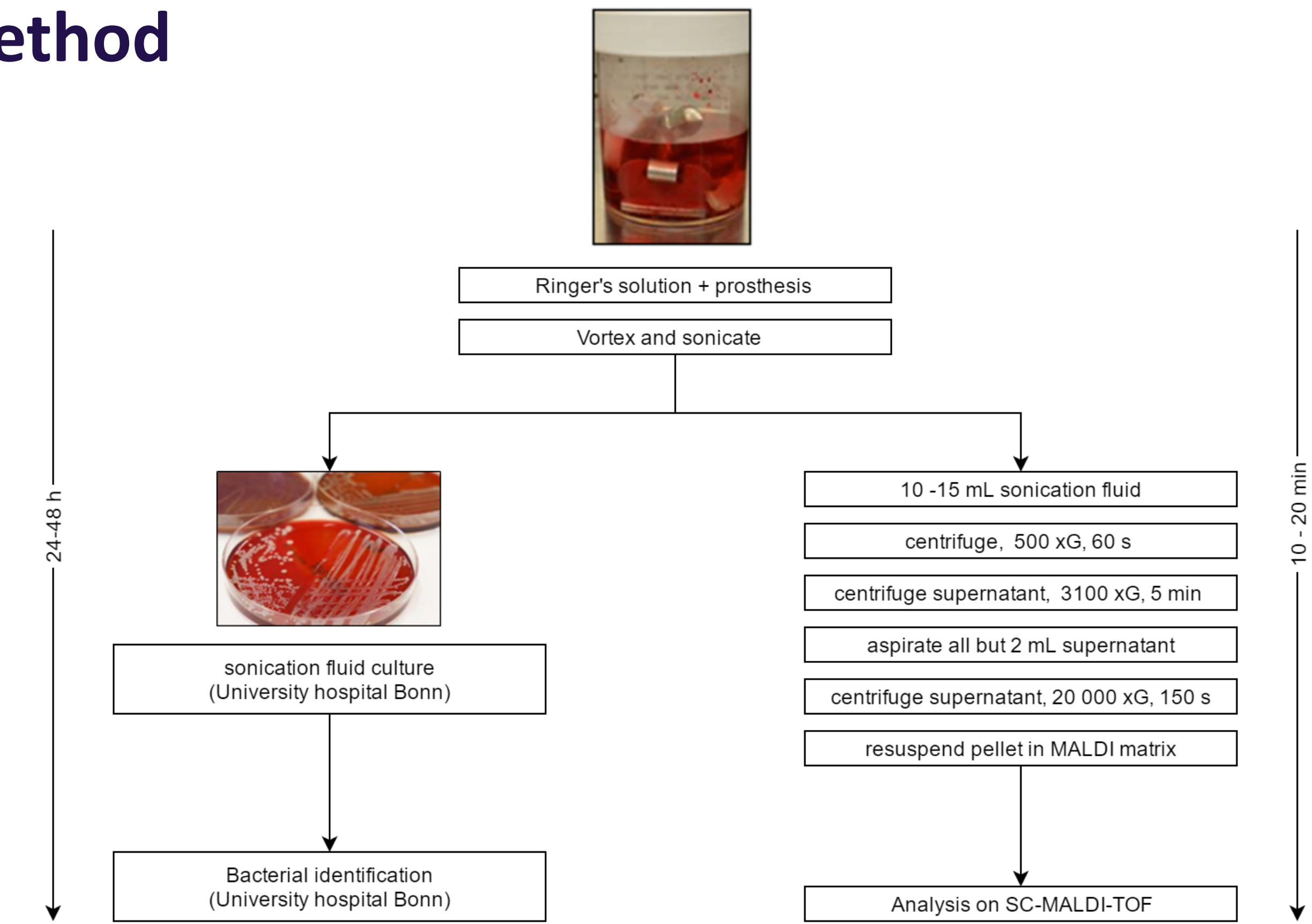


Fig. 1. Sample processing method for SC-MALDI-TOF and culture-based identification

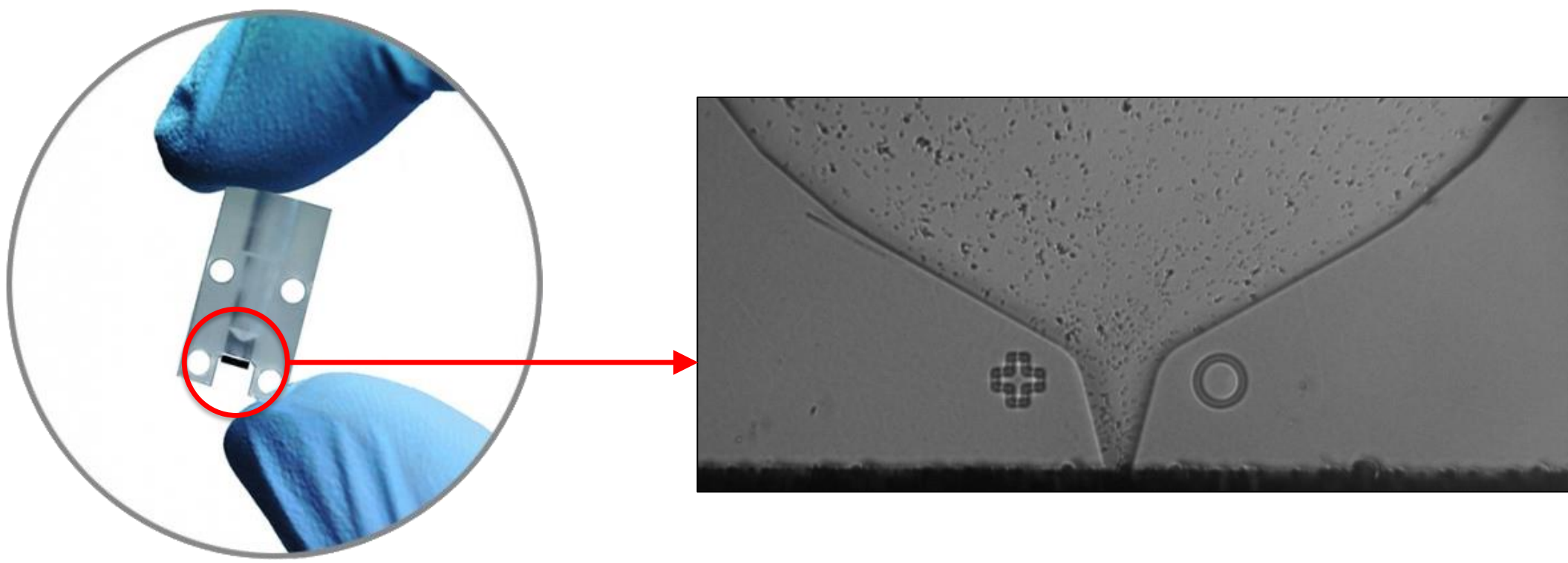


Fig. 2. SC-MALDI-TOF; Each cell is presented in a separate particle to the MALDI-TOF, resulting in a mass spectrum for every cell. Image courtesy of cytena.

Results

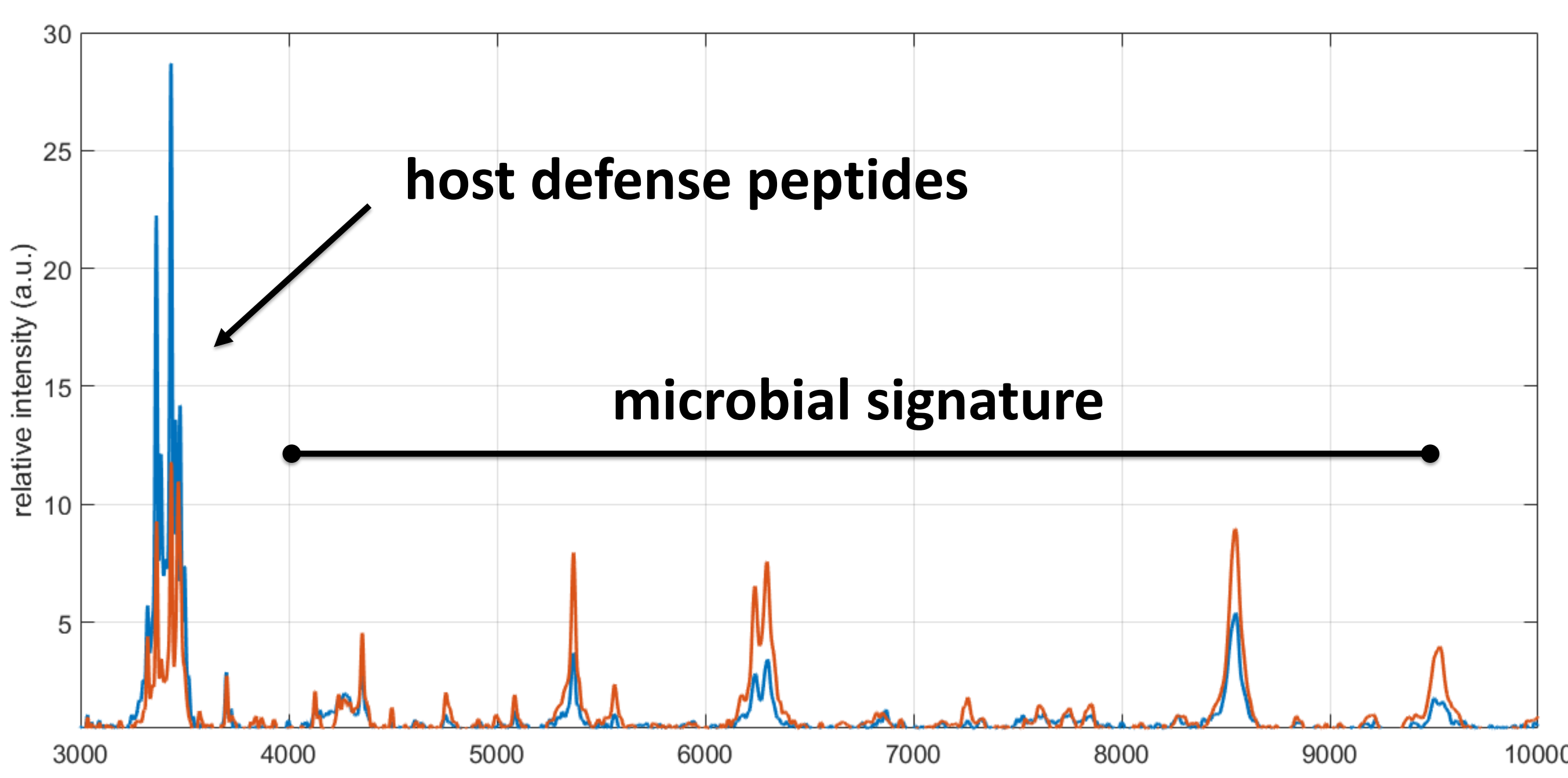


Fig. 3. Mass spectra of *S. epidermidis* obtained directly from sonication fluid. In addition to the microbial signature (4 – 10 kDa), several peaks with masses corresponding to α -defensins 1-4 are observed (resp. 3373, 3445, 3488, and 3716 Da). The intensity of the α -defensins is inversely correlated with intensity of the microbial signature.

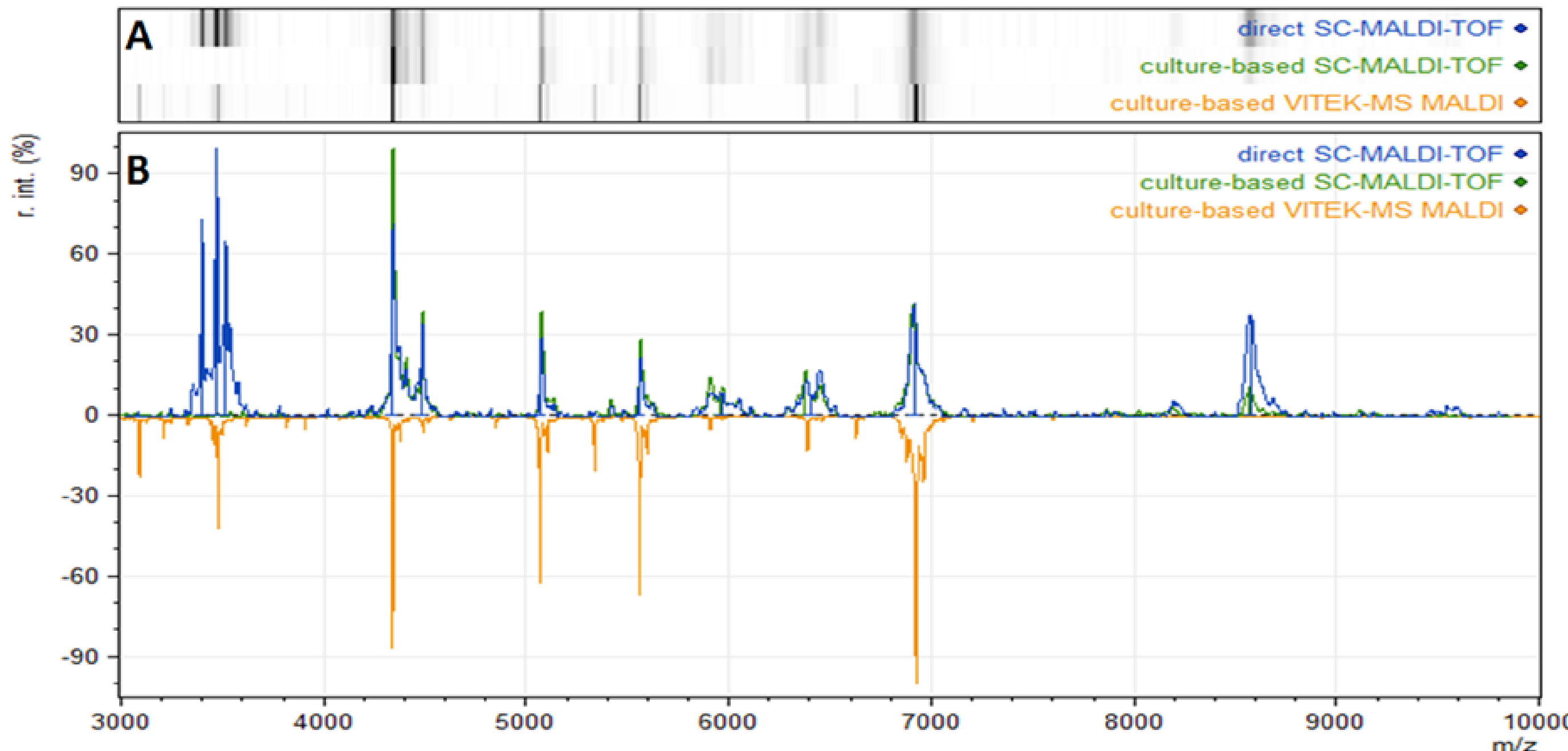


Fig. 4. Pseudo-gel (A) and mass spectrum (B) of *S. aureus* directly from sonication fluid (blue) and culture-based single-cell MALDI-TOF (green) and conventional MALDI-TOF (yellow). The α -defensins are solely observable using direct MALDI-TOF.

Table. 1. Summarized results of direct SC-MALDI-TOF MS of 27 clinical sonication fluid aliquots

| Reference identification | presence of α -defensins (%) | microbial signature (%) | signal suppression* (%) |
|-----------------------------|-------------------------------------|-------------------------|-------------------------|
| <i>E. coli</i> (n=1) | 1 (100) | 1 (100) | 1 (100) |
| <i>Enterococci</i> (n=3) | 3 (100) | 2 (67) | 3 (100) |
| <i>P. acnes</i> (n=3) | 3 (100) | 3 (100) | 2 (67) |
| <i>S. aureus</i> (n=2) | 2 (100) | 2 (100) | 1 (50) |
| <i>S. epidermidis</i> (n=6) | 6 (100) | 4 (67) | 5 (83) |
| Culture negative (n=12) | 11 (92) | 4 (33) | 7 (58) |

*mass spectra with high intensity α -defensin peaks associated with a reduced intensity of the microbial signature

Conclusions

This study shows the technical feasibility of a new MALDI method, that can analyse sonicates within minutes after collection during surgery by avoiding the time-consuming and potentially false negative culture step:

- Bacteria can be observed directly from sonication fluid: in 80% of the positive culture samples and in 33% negative culture samples a microbial signature was observed.
- Mass spectra obtained using direct SC-MALDI-TOF do contain extra information about host-pathogen interaction: peaks corresponding to α -defensins were found in 100% of the positive culture samples and 92% of the negative culture samples.
- 80% of the mass spectra obtained from positive sonication aliquots showed intense α -defensin peaks in combination with a (severely) reduced microbial spectrum or no microbial spectrum at all. For 58 % of the negative sonication aliquots this was the case

Discussion

A relatively high number of microbial signatures were observed from culture negative sonicates. Non-culturable bacteria cannot be analysed by culture-dependent MALDI-TOF, while bacteria originating from biofilms, as is the case in most PJIs, have been found to exist in a viable but non-culturable state¹. In addition, bacteria could be severely injured by host defence peptides (e.g. α -defensins) or preoperative antibiotic administration, and therefore non-viable or non-culturable.

The high amount of α -defensins in some samples implies the presence of an infecting pathogen, although in some cases no bacterial signature was found. α -defensin has received a lot of attention in the recent years as biomarker for PJI and a recent developed assay has shown to be an useful tool to confirm the absence of PJI².

The intensity of the α -defensins is in most mass spectra inversely correlated with intensity of the microbial signature. The cationic α -defensins have a relatively low ionization potential and are therefore likely suppress other proteins in MALDI-TOF. This phenomenon has been described in literature before in the case of direct MALDI-TOF of urine samples³.

Acknowledgments

Special thanks to Dr. S.A.J. Zaat, medical microbiology, Academic Medical Center Amsterdam. This research will continue in Eurostars project SIBaGEM (ID 10 257) in cooperation with Academic Medical Center Amsterdam, Albert Ludwigs Universität Freiburg and cytena.

(1) Pasquaroli, S., Zandri, G., Vignaroli, C., Vuotto, C., Donelli, G., & Biavasco, F. (2013). Antibiotic pressure can induce the viable but non-culturable state in *Staphylococcus aureus* growing in biofilms. *Journal of Antimicrobial Chemotherapy*, 68(8), 1812-1817.

(2) Kasperek, M. F., Kasperek, M., Boettner, F., Faschingbauer, M., Hahne, J., & Dominkus, M. (2016). Intraoperative diagnosis of periprosthetic joint infection using a novel alpha-defensin lateral flow assay. *The Journal of Arthroplasty*, 31(12), 2871-2874.

(3) Köhling, H. L., Bittner, A., Müller, K. D., Buer, J., Becker, M., Rübben, H., Rettenmeier, A.W., Mosel, F. (2012). Direct identification of bacteria in urine samples by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry and relevance of defensins as interfering factors. *J Med Microbiol*, 61(Pt 3), 339-344.